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ISOLATION AND STRUCTURE OF CITREOPYRONE, A METABOLITE OF PENICILLIUM CITREO-VIRIDE BIOURGE

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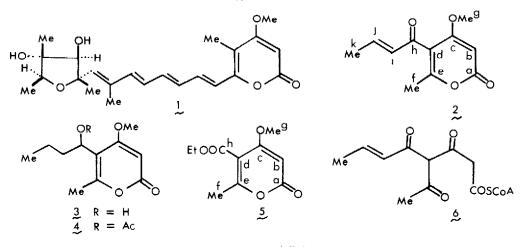
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<u>Summary</u>. Citreopyrone isolated from the mycelium of <u>Penicillium citreo-viride</u> B. has been found to be 5-crotonoyl-4-methoxy-6-methyl-2-pyrone. This is the first naturally occurring pyrone with an acyl group at C<sub>5</sub>-position.

Of many substances produced by mould and fungus, pyrones which are regarded as masked  $\ell$ -polyketo carboxylic acids are quite interesting from view points of their chemical reactivity<sup>1</sup> as well as of biological activity.<sup>2</sup> Citreoviridin, a toxic yellow metabolite of <u>P</u>. <u>citreo-viride</u> B., was first isolated by Hirata,<sup>3</sup> and its structure was also elucidated to be <u>1</u>.<sup>4</sup> In the present paper, we wish to describe the isolation and structure of a new pyrone (<u>2</u>), called citreopyrone. This is the first naturally occurring pyrone with an acyl group at C<sub>5</sub>-position.

Polished rice (250 g) in deionized water (800 m1) was allowed to stand at room temperature for 30 mln, then cooked using an electric rice cooker (99 °C, 17 min) and transfered into an Erlenmyer flask (3 1), which was pasteurized (120 °C, 20 min at 2 atom), then inoculated with a suspension of mycelium of <u>P. citreo-viride</u> B. (IFO 6200) in a sterilized water and incubated stationarily at 24 °C for 32 days. The yellow rice thus obtained was extracted with AcOEt and then with acetone. The combined extracts were concentrated under reduced pressure, and then separated by a combination of column chromatography (Mallinckrodt, 100 mesh; AcOEt)<sup>5</sup> and repeated preparative TLC [Kieselgel PF<sub>254</sub>; CHCl<sub>3</sub> - AcOEt (1 · 1)] to afford citreopyrone (2) (17 mg) as colorless columns [mp 109 - 109.5 °C;  $c_{11}H_{12}O_4$  (m/e 208(M<sup>+</sup>))]. The IR and UV spectra [ $\mathcal{Y}_{max}$ (Nujol) 1735, 1635 and 1565 cm<sup>-1</sup>;  $\lambda_{max}$ (MeOH) 273 nm (£ 7300)] of 2 indicate the presence of an  $\mathscr{L}$ -pyrone, which has three substituents: Me [ $\mathcal{E}$ 2.18(3H, s)], MeO [ $\mathcal{E}$ 3.80(3H, s)] and MeCH<sup>E</sup>\_{ECHCO} [ $\mathcal{Y}_{max}$ (Nujol) 1680 and 1610 cm<sup>-1</sup>;  $\mathcal{E}$ 1.97(3H, dd, J= 7, 1.5Hz), 6.26(1H, dq, J= 16, 1.5Hz) and 6.76(1H, dq, J= 16, 7Hz)].<sup>6</sup> Particularly, the presence of the crotonoyl group was confirmed by  $NaBH_4$  reduction of 2 in THF (0 °C, 50 min) leading to the formation of the corresponding tetrahydro compound (3) as a colorless oil  $[C_{11}H_{16}O_4 \text{ (m/e 212(M^+)); } \gamma_{max}(\text{film}) 3400 \text{ cm}^{-1}; \$0.93(3H, t, J= 7Hz), 1.1 - 1.9(4H, t)$ complex) and 4.61(1H, t, J= 7Hz)]. The methine triplet at §4.61 in 3 was shifted to §5.76 in the  $^{1}$ H NMR spectrum of the corresponding acetate (4) which was readily obtained on acetylation of the former with  $Ac_20$  - pyridine.

Finally, the positions of each substituent on the  $\mathcal{L}$ -pyrone ring were based on a comparison of  ${}^{13}$ C NMR spectra between 2 and 5 [2: \$162.7(s) (C<sup>a</sup>), 87.5(d) (C<sup>b</sup>), 168.3(s) (C<sup>c</sup>), 113.7(s) (C<sup>d</sup>), 161.0(s) ( $C^{e}$ ), 18.4(q) ( $C^{f}$ ), 56.2(q) ( $C^{g}$ ), 190.0(s) ( $C^{h}$ ), 132.7(d) ( $C^{i}$ ), 146.4(d) ( $C^{j}$ ) and 18.2 (q) ( $C^{k}$ ). 5: **b**163.1(s) or 163.3(s) ( $C^{a}$ ), 87.5(d) ( $C^{b}$ ), 167.9(s) ( $C^{c}$ ), 108.8(s) ( $C^{d}$ ), 162.1(s)  $(C^{e})$ , 18.6(q)  $(C^{f})$ , 56.5(q)  $(C^{g})$ , 163.3(s) or 163.1(s)  $(C^{h})$ , 61.6(t) and 14.1(q) (Et)].<sup>7</sup> In the <sup>1</sup>H NMR spectra, furthermore, both 2 and 5 have a singlet assignable to  $C_3$ -H at §5.45 and 5.44, respectively. Biogenetically, citreopyrone (2) may be derived from a plausible intermediate (6).



## References and Notes

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  After elution of 2, citreoviridin (1) (237 mg) was obtained.
  I. H and 13C NMR spectra of these pyrones were measured on a JEOL PS-100 or FX-100 NMR spectro-meter using CDCl3 as the solvent and TMS as the internal standard.
  The chemical shifts of each corresponding signal in both 2 and 5 are quite similar to each other except for the chemical shifts assignable to C<sup>d</sup> (\$113.7 in 2; \$108.8 in 5). Clearly, this difference is explained well by remarkable differences of the two different types of C0 group in their effects on the C atom directly attached to them. group in their effects on the C atom directly attached to them.

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